



OPEN ACCESS

EDITED BY

Francesco Bussu,
University of Sassari, Italy

REVIEWED BY

Jenni Virtanen,
University of Helsinki, Finland
Davide Rizzo,
University of Sassari, Italy

*CORRESPONDENCE

Tina Uršič

✉ tina.ursic@mf.uni-lj.si

SPECIALTY SECTION

This article was submitted to
Infectious Diseases: Epidemiology and
Prevention,
a section of the journal
Frontiers in Public Health

RECEIVED 12 October 2022

ACCEPTED 06 March 2023

PUBLISHED 23 March 2023

CITATION

Uršič T, Kogoj R, Šikonja J, Jevšnik Virant M and
Petrovec M (2023) A simplified nasopharyngeal
swab collection procedure for minimizing
patient discomfort while retaining sample
quality. *Front. Public Health* 11:1066934.
doi: 10.3389/fpubh.2023.1066934

COPYRIGHT

© 2023 Uršič, Kogoj, Šikonja, Jevšnik Virant and
Petrovec. This is an open-access article
distributed under the terms of the [Creative
Commons Attribution License \(CC BY\)](#). The use,
distribution or reproduction in other forums is
permitted, provided the original author(s) and
the copyright owner(s) are credited and that
the original publication in this journal is cited, in
accordance with accepted academic practice.
No use, distribution or reproduction is
permitted which does not comply with these
terms.

A simplified nasopharyngeal swab collection procedure for minimizing patient discomfort while retaining sample quality

Tina Uršič*, Rok Kogoj, Jaka Šikonja, Monika Jevšnik Virant and
Miroslav Petrovec

Institute of Microbiology and Immunology, Faculty of Medicine, University of Ljubljana, Ljubljana, Slovenia

A nasopharyngeal swab (NPS) is the most frequently collected sample type when molecular diagnosis of respiratory viruses, including SARS CoV-2, is required. An optimal collection technique would provide sufficient sample quality for the diagnostic process and would minimize the discomfort felt by the patient. This study compares a simplified NPS collection procedure with only one rotation of the swab to a more standard procedure with five rotations. Swabs were collected from 76 healthy volunteers by the same healthcare professional on 2 consecutive days at a similar hour to minimize variability. The number of Ubiquitin C copy number per sample was measured by real-time quantitative PCR and patient discomfort was assessed by questionnaire. No statistically significant difference ($p = 0.15$) was observed in the Ubiquitin C copy number per sample between a NPS collected with one rotation ($5.2 \pm 0.6 \log UBC$ number copies/sample) or five rotations ($5.3 \pm 0.5 \log UBC$ number copies/sample). However, a statistically significant difference was observed in discomfort between these two procedures, the second being much more uncomfortable. Additional analysis of the results showed a weak correlation between discomfort and the number of human cells recovered (Spearman's $\rho = 0.202$) and greater discomfort in younger people. The results of this study show that a NPS collected with one slow rotation has the same quality as a NPS collected with five rotations. However, the collection time is shorter and, most importantly, less unpleasant for patients.

KEYWORDS

nasopharyngeal swab, collection procedure, sample quality, discomfort score, Ubiquitin C

1. Introduction

A nasopharyngeal swab (NPS) is the most commonly collected specimen when laboratory diagnosis of respiratory viral infections, including COVID-19, is required. As an appropriate NPS collection technique is critical for reliable results and for minimizing patient discomfort during the procedure (1), so is the proper use of personal protective equipment to minimize the risk of health care professionals' infection (2). Current recommendations for obtaining NPS are not uniform. The World Health Organization (WHO) recommends leaving the swab in place for a few seconds after reaching the nasopharynx before withdrawing it (3), the Center for Disease Control and Prevention (CDC) recommends "gently rubbing and rolling" the swab after reaching the nasopharynx and leaving it in place for several seconds before withdrawing it (4). To further address these issues and provide additional guidance, Marty et al. published an article with practical

instructions and a descriptive video in *The New England Journal of Medicine* early in the pandemic (5). Nevertheless, we wondered if the procedure could be further changed to reduce the proportion of patients that find it frustrating or even painful, and simplified without compromising the quality of the sample.

The aim of this study was to investigate whether the NPS collection could be simplified to only one rotation with no waiting period in the nasopharynx yet retain the sample quality but decrease the discomfort felt by the participants.

2. Methods

2.1. Study participants, sample collection, and processing

In this study, we collected 3 NPS samples from 76 healthy adults without any respiratory symptoms over the course of 7 days. A single healthcare provider collected all NPSs to minimize differences in the swab collection procedure. We used the CITOSWAB VTM (Nal Von Minden, Regensburg, Germany) collection kit containing 3 ml of viral transport medium. NPS were collected on 2 consecutive days at around the same time. On the 1st day, sample A was collected with one rotation and sample B with five rotations one after another with enough time in between to allow the participant to recover from the discomfort experienced. On the 2nd day, only a NPS with five rotations (sample C) was collected. The detailed method for sample A collection was:

- Gentle insertion of the swab through the nostril;
- Simultaneous pushing and rotating of the swab 5–7 cm to the nasopharynx and immediate removal with simultaneous rotation during withdrawal amounting to one full rotation;

Samples B and C were collected by following previously published recommendations (5).

Study participants were asked to rate their discomfort immediately after collection on a scale from 1 representing absence or no discomfort, linearly to 10 representing unbearable discomfort.

Samples were blinded and processed within 3 h after sample collection. Total nucleic acids (NA) were extracted using a MagNA Pure Compact instrument (Roche, Mannheim, Germany). The swab collection quality was assessed by quantifying the amount of human UBC gene copies per sample (6) by using LightMix[®] Kit SARS-CoV-2 E+N UBC (TibMolbiol, Berlin, Germany) with an in-house UBC quantification standard. The result was finally expressed as log [UBC copies/sample] taking into account volume calculations, 200 μ l of sample volume and 100 μ l of elution volume.

2.2. Statistical methods

Differences in sample quality expressed as log [UBC copies/sample] were evaluated with parametric tests. Discomfort scores, classified as ordinal data, were compared with non-parametric tests. Therefore, for in-group and between-group comparisons, a paired *t*-test (or Wilcoxon signed-rank test) and

independent samples *t*-test (Mann–Whitney *U*-test) were used, respectively. The strength of the association between discomfort scores and log [UBC copies/sample] was assessed using Spearman's rank-order correlation. A value of $p \leq 0.05$ was considered statistically significant in all tests. Continuous data, such as log [UBC copies/sample], are presented as mean \pm standard deviation, whereas ordinal data (discomfort score) are presented as median (first–third quartile).

3. Results

The mean age of the 76 study participants was 36 ± 11 years, and 56 (74%) were female. The observed median discomfort score (DS) for sample A was 3 (first-third quartile; 2–5), whereas for sample B it was 6 (first-third quartile; 4.25–7) and for sample C it was also 6 (first-third quartile; 4–7), as shown in Figure 1A. The DS of sample A was significantly lower when compared to samples B and C ($p < 0.001$). No statistically significant difference was observed between samples B and C ($p = 0.59$).

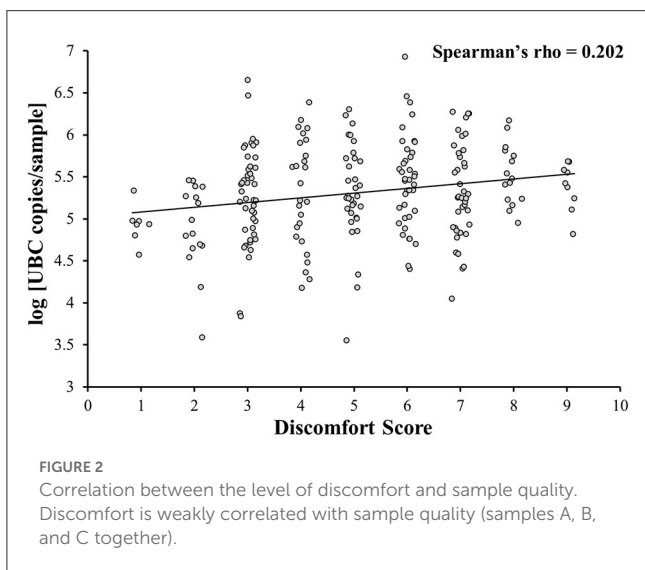
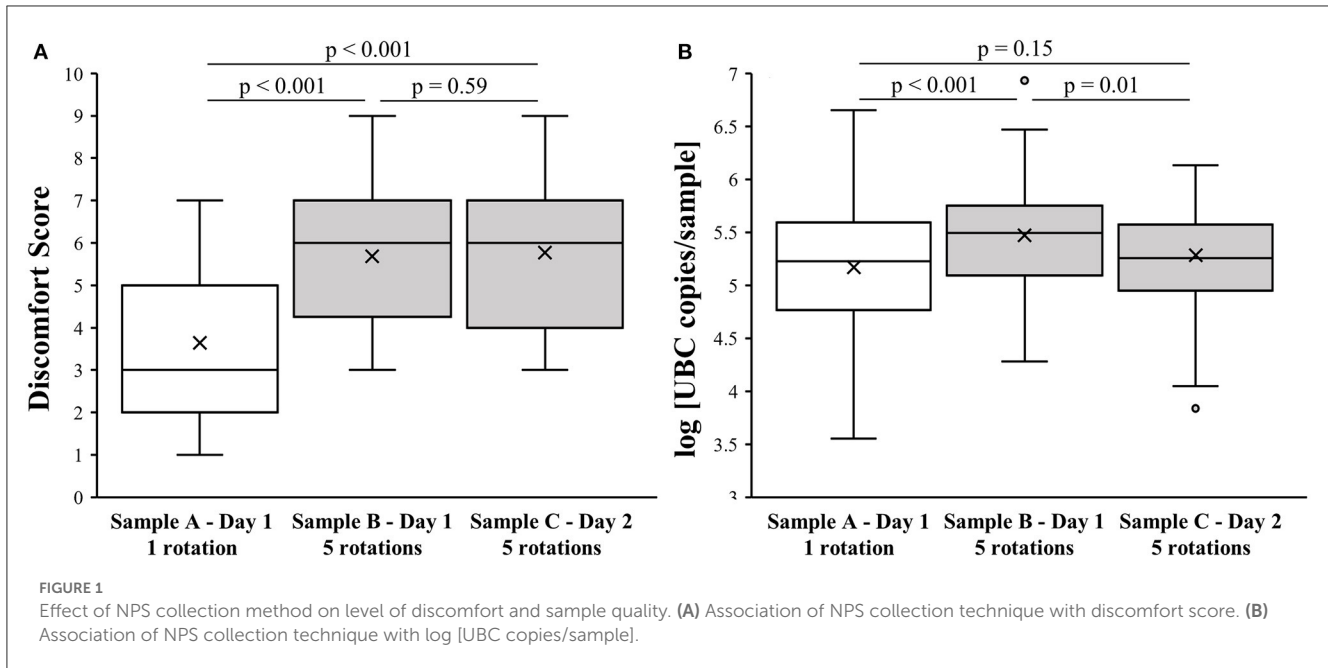
There was no significant difference in the log [UBC copies/sample] between samples A and C (5.2 ± 0.6 vs. 5.3 ± 0.5 ; $p = 0.15$). However, sample B had higher log [UBC copies/sample] values (5.5 ± 0.5) compared to samples A and C ($p < 0.001$ and $p = 0.01$, respectively), as presented in Figure 1B.

Although younger participants (≤ 35 years) reported significantly higher DS in comparison to older participants (sample A, median: 4 vs. 3, $p = 0.007$; sample B, median: 6 vs. 5, $p = 0.02$; sample C, median: 7 vs. 5.5, $p = 0.056$), the two age groups did not differ in mean log [UBC copies/sample]. Moreover, comparable DS were observed between men and women (sample A, median: 3.5 vs. 3, $p = 0.843$; sample B, median: 5.5 vs. 6, $p = 0.815$; sample C, median: 6 vs. 6, $p = 0.728$) as were also log [UBC copies/sample] $p > 0.1$.

A weak positive correlation between DS and log [UBC copies/sample] was observed (Spearman's rho = 0.202; $p = 0.002$; Figure 2). Additional analysis on samples A, B and C together showed that NPS collected with minimal subject discomfort (DS < first quartile) had a lower log [UBC copies/sample] than NPS collected with high subject discomfort (DS > third quartile; 4.9 ± 0.4 vs. 5.5 ± 0.3 ; $p < 0.001$). When looking separately, only sample A showed that NPS collected with minimal subject discomfort (DS < first quartile) had a lower log [UBC copies/sample] than NPS collected with high subject discomfort (4.9 ± 0.2 vs. 5.7 ± 0.4 ; $p < 0.001$) and no link between DS and log [UBC copies/sample] was observed in samples B (5.4 ± 0.6 vs. 5.3 ± 0.4 ; $p = 0.42$) and C (5.2 ± 0.5 vs. 5.4 ± 0.3 ; $p = 0.22$).

4. Discussion

The results of this study show no statistically significant difference in the UBC gene copies/sample between NPS collected with one or five rotations (sample A vs. sample C). These results are also supported by the study from Kinloch et al. (7). They similarly found no difference in DNA/RNA recovery between an “in-out” NPS and a NPS with 10 s of rotation. Our study found a significantly higher DS of NPS with five rotations compared to one rotation



NPS. Interestingly, in contrast to our results, Kinloch et al. found no differences in discomfort between the procedures they used (7).

Despite all efforts for uniformity of the collection procedure, differences in log [UBC copies/sample] values were still found between study participants, which may indicate the diversity of the nasal mucosa and anatomy of the individuals (7, 8). In the study by Kinloch et al., RPP30 and RNase p-values also varied markedly regardless of the swab collection technique (7).

Although only a weak correlation between DS and log [UBC copies/sample] was observed between the participants that reported almost no discomfort and those that reported the highest DS, the difference between log [UBC copies/sample] reached a statistically significant value. This observation theoretically means that samples with higher human cell counts would have an impact on specific target (pathogen) detection sensitivity, however due to the study design such a conclusion can't be drawn. Similar results were also

observed by Kinloch et al., who showed statistically significant higher DNA/RNA recovery together with higher DS in Asian participants of their study population (7, 8).

Finally, sample B was initially collected because we wanted to gain insight into the DS and quality of the sample if one sample is collected right after another. Contrary to expectations, we found UBC copies/sample in sample B higher than in sample A and even sample C, however, the difference was rather small. We believe that minor trauma to the nasal pseudostratified ciliated epithelium may have occurred with the collection of sample A, which could explain the higher UBC copies/sample in sample B.

A few limitations of the study must be kept in mind when interpreting the results. Firstly, the study population was rather small, due to difficulties of acquiring willing participants after 2 years of frequent NPs collections for SARS-CoV-2 screening. Secondly, only adults were included in the study, therefore the results do not necessarily apply to children, especially young children who do not yet understand why NPs is being collected and are usually already under stress at the collection site which makes them hindered to give an rational DS score. Thirdly, the study was performed on healthy individuals and different results might occur in individuals with acute respiratory infection, who usually secrete increased amounts of mucus and cellular material. Finally, if NPSs were collected by multiple healthcare professionals, although with the same protocol, additional variance might have been observed in both, quality and DS. Future studies should address these questions.

5. Conclusion

In conclusion, this study shows that a simplified NPS collection procedure has the same recovery of human DNA as a NPS collected with five rotations. However, the collection time is shorter and the experience is less unpleasant for patients. We firmly believe that the NPS collection procedure could be improved globally (3, 4, 7).

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Ethics statement

The studies involving human participants were reviewed and approved by National Medical Ethics Committee of Slovenia (No. 0120-281/2022/3). The patients/participants provided their written informed consent to participate in this study.

Author contributions

TU: conceptualization, methodology, visualization, and writing—original draft preparation. RK: writing—reviewing and editing, supervision, and reviewing. JŠ: software, data curation, and writing—original draft preparation. MJ: investigation, validation, and reviewing. MP: supervision, reviewing, and editing. All authors contributed to the article and approved the submitted version.

Funding

This study was supported by the Slovenian Research Agency (Research Program P3-0083).

References

- Pondaven-Letourmy S, Alvin F, Boumghit Y, Simon F. How to perform a nasopharyngeal swab in adults and children in the COVID-19 era. *Eur Ann Otorhinolaryngol Head Neck Dis.* (2020) 137:325–7. doi: 10.1016/j.anorl.2020.06.001
- Piras A, Rizzo D, Longoni E, Turra N, Urru S, Saba PP, et al. Nasopharyngeal swab collection in the suspicion of Covid-19. *Am J Otolaryngol.* (2020) 41:102551. doi: 10.1016/j.amjoto.2020.102551
- Organization WH. *Interim Guidance 19 March 2020. Laboratory Testing for Coronavirus Disease (COVID-19) in Suspected Human Cases.* Geneva:WHO (2020).
- Center for Disease Control and Prevention. *Interim Guidelines for Collecting and Handling of Clinical Specimens for COVID-19 testing.* (2022). Available online at: <https://www.cdc.gov/coronavirus/2019-ncov/lab/guidelines-clinical-specimens.html>
- Marty FM, Chen K, Verrill KA. How to obtain a nasopharyngeal swab specimen. *N Engl J Med.* (2020) 382:e76. doi: 10.1056/NEJMvem2010260
- Chua SL, See Too WC, Khoo BY, Few LL. UBC and YWHAZ as suitable reference genes for accurate normalisation of gene expression using MCF7, HCT116 and HepG2 cell lines. *Cytotechnology.* (2011) 63:645–54. doi: 10.1007/s10616-011-9383-4
- Kinloch NN, Shahid A, Ritchie G, Dong W, Lawson T, Montaner JSG, et al. Evaluation of nasopharyngeal swab collection techniques for nucleic acid recovery and participant experience: Recommendations for COVID-19 diagnostics. *Open Forum Infect Dis.* (2020) 7:ofaa488. doi: 10.1093/ofid/ofaa488
- Morgan NJ, MacGregor FB, Birchall MA, Lund VJ, Sittampalam Y. Racial differences in nasal fossa dimensions determined by acoustic rhinometry. *Rhinology.* (1995) 33:224–8.

Acknowledgments

The authors would like to express their gratitude to all the participants that were willing to participate in this study. Many thanks to Sabina Beci, Gašper Grubelnik, Kaja Kotnik, Manca Luštrek, Špela Pleh, Katja Potočnik, and Zala Prestor for their excellent laboratory work. We would also like to express our gratitude to Gregory A. Storch from the Washington University School of Medicine in St. Louis, Missouri, USA for the valuable manuscript review.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.